Vibralactone: A Lipase Inhibitor with an Unusual Fused β -Lactone Produced by Cultures of the Basidiomycete Boreostereum vibrans

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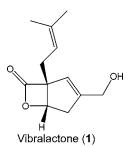
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ABSTRACT



The structure and absolute configuration of vibralactone (1) from the cultures of the Basidiomycete Boreostereum vibrans were established by spectroscopic methods and computational methods. Vibralactone, an unusual fused β -lactone-type metabolite, was found to inhibit pancreatic lipase with an IC₅₀ of 0.4 μ g/mL.

Lipid metabolism normally keeps a delicate balance between synthesis and degradation. When the balance is upset, hyperlipidemia may occur, which in turn can cause atherosclerosis, hypertension, diabetes, etc.¹ Modulators of lipid metabolism are expected to be useful in controlling these disorders. Obesity and hypercholesterolemia are to a relevant degree related to high nutritional fat intake. The key enzyme of dietary triglyceride absorption is pancreatic lipase. Inhibition of pancreatic lipase may therefore result in inhibition

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of fat absorption. Orlistat, a specific pancreatic lipase, is clinically used for preventing obesity and hyperlipidemia.² Because of the biodiversity, chemodiversity, and unexplored resources of the fungal kindom, secondary metabolites from macrofungi in China were investigated,3-6 and a compound called vibralactone was isolated from the culture broth of the polypore Boreostereum vibrans (Berk & M. A. Curtis; Davydkina & Bondartseva (Aphyllophorales)). It shows inhibition of pancreatic lipase with an IC₅₀ of 0.4 μ g/mL. In

- (3) Liu, J. K. Chem. Rev. 2006, 106, 2209.
- (4) Liu, J. K. Chem. Rev. 2005, 105, 2723.

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⁽¹⁾ Borgstrom, B. In Exocrine Pancrease, Pathology and Diseases; Go, V. L., et al., Eds.; Raven Press: New York, 1981; pp 361-373.

⁽²⁾ Hill, J. Q.; Hauptman, J.; Anderson, J. W.; Fujioka, K.; O'Neil, P. M.; Smith, D. K.; Zavoral, J. H. Am. J. Clin. Nutr. **1999**, 69, 1108.

 ⁽⁵⁾ Qin, X. D.; Dong, Z. J.; Liu, J. K.; Yang, L. M.; Wang, R. R.; Zheng,
 Y. T.; Lu, Y.; Wu, Y. S.; Zheng, Q. T. *Helv. Chim. Acta* 2006, *89*, 127.
 (6) Liu, J. K. *Heterocycles* 2002, *57*, 157.

previous investigations, a series of sterpurane-type sesquiterpenoids have been isolated from the closely related genus *Stereum*, and these compounds exhibited broad biological activities, including phytotoxic, antibiotic, and causative activities of the silver leaf disease.^{7–10} In this communication, we report on the structural elucidation and assignment of the absolute configuration of vibralactone, an unusual β -lactone-type metabolite.

Vibralactone was obtained as a colorless oil, $[\alpha]_D^{26} =$ -135.1 (c = 0.517, CHCl₃). It exhibited a quasimolecular ion peak at m/z 231 corresponding to $[M + Na]^+$ in the positive ESI-MS, and HR-ESI-MS analysis provided the molecular formula $C_{12}H_{16}O_3$ (calcd for $[M + Na]^+ m/z$ 231.1004, found 231.0997). The IR spectrum of vibralactone exhibited carbonyl and hydroxyl group absorptions at 1816 and 3423 cm⁻¹, respectively. Its ¹³C NMR spectrum showed 12 resolved peaks corresponding to 12 C-atoms, which were classified into 2 Me, 2 CH₂, 1 OCH₂, 1 OCH, 2 sp² CH, 2 sp² C, 1 sp³ C, and 1 C=O groups by analysis of the DEPT spectra. An analysis of the ¹H NMR spectrum indicated the presence of two olefinic protons at $\delta = 5.60$ (1H, s) and 5.11 ppm (1H, t, J = 7.3 Hz), in addition to two isolated methyl groups at $\delta = 1.71$ (3H, s) and 1.62 ppm (3H, s). The ¹³C NMR spectrum showed four olefinic carbon resonances at $\delta = 146.6$ (s), 136.0 (s), 122.4 (d), and 117.2 ppm (d) as well as a carbonyl signal at $\delta = 173.0$ ppm (s).

The connectivity of the protons and C-atoms was established by the ¹H,¹³C HSQC spectrum. The cross-peaks between H-2 and H-4/H-13, H-9 and H-8/H-11/H-12, H-5 and H-4, and H-13 and H-4 were observed in the ¹H,¹H COSY spectrum. It allowed establishment of two H-atom systems, one at C-8 through C-11 and C-12 and the other at C-2 through C-13 and C-5. ¹³C, ¹H long-range couplings (³J) observed in the HMBC experiments gave the following correlations from H-2 to C-4, C-5, and C-13, from H-9 to C-1, C-11, and C-12, from H-5 to C-2, C-3, C-7, and C-8, from H-13 to C-2 and C-4, from H-4 to C-2, from H-8 to C-2, C-5, and C-10, from H-12 to C-9 and C-11, and from H-11 to C-9 and C-12. In addition, the HMBC spectrum showed unusual correlations across five bonds between H-13 and C-7 as well as between H-13 and C-8. It also happened in the same compound, and we observed the correlations across five bonds between H-11, H-12, and C-1. The presence of the substructure from C-1 through C-8, C-9, C-10, C-11, and C-12 in vibralactone is doubtless. The presence of a double bond in the five bonds of this unique structure could be the reason. By combining all this evidence and data, we were able to assign planar structure 1 to vibralactone (Table 1, Figure 1). It explains the characteristic IR absorption at 1816 cm⁻¹ for the β -lactone as well as the singlet for the CH₂OH group at $\delta_{\rm H} = 4.22$ ppm ($\delta_{\rm C} = 61.3$ ppm) in the ¹H NMR spectrum.

The sharp singlet for an olefinic proton at $\delta_{\rm H} = 5.60$ ppm attributed to H-2 in the ¹H NMR spectrum suggested that

Table 1.		NMR Spectral Data for Vibralactone (1) in CDCl ₃		
	¹³ C	$^{1}\mathrm{H}$	¹ H, ¹ H COSY	HMBC
1	75.1			
2	122.4	5.60 (1H, s)	H-4, 13	C-1, 3, 4, 5, 13
3	146.6			
4	37.3	$2.77~(1\mathrm{H},\mathrm{dd},18.8,4.3)$	H-2, 5, 13	C-2, 3, 5
		2.71 (1H, d, 18.8)		
5	78.5	4.79 (1H, d, 4.3)	H-4	C-2, 3, 4, 7, 8
7	173.0			
8	27.6	$2.60(1\mathrm{H},\mathrm{dd},15.1,7.3)$	H-8b, 9	C-1, 2, 5, 9, 10
		$2.41(1\mathrm{H},\mathrm{dd},15.1,7.3)$	H-8a, 9	
9	117.2	5.11 (1H, t, 7.3)	H-8, 11, 12	C-1, 11, 12
10	136.0			
11	18.0	1.62(3H, s)	H-9	C-1, 9, 10, 12
12	25.8	1.71 (3H, s)	H-9	C-1, 9, 10, 11
13	61.3	4.22 (2H, s)	H-4	C-1, 2, 3, 4, 7, 8

the CH₂OH group was attached to C-3. This was confirmed by the correlations between H-4 and H-13 in the 1 H, ¹H COSY spectrum and between H-2 and H-8 in the HMBC. Further evidence for this is the presence of cross-peaks between H-2 and H-8 and H-13 in the ROESY.

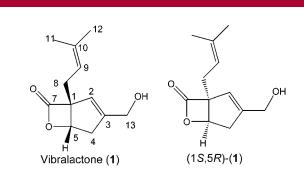


Figure 1. Structure of vibralactone (1).

A literature search indicated that all 6-oxy-bicyclo[3.2.0]-hept-2-en-7-ones reported so far are cis-fused. The transfused species can be excluded due to the instability of the lactone group caused by the high strain energy of this system.^{11–14} The cis stereochemistry of vibralactone is supported by computation of the ¹³C NMR shifts for the cis and trans stereoisomers (see Supporting Information).

For the assignment of absolute configuration, the optical rotation values for vibralactone (1) and its enantiomer (1S,5R)-1 were calculated. The first use of modern HF calculations for optical rotations was reported by Polavarapu

⁽⁷⁾ Mellows, G.; Mantle, P. G.; Feline, T. C.; Williams, D. J. *Phytochemistry* **1973**, *12*, 2717.

⁽⁸⁾ Ayer, W. A.; Saeedi-Ghomi, M. H. Can. J. Chem. 1981, 59, 2536.
(9) Ayer, W. A.; Saeedi-Ghomi, M. H. Tetrahedron Lett. 1981, 22, 2071.

⁽¹⁰⁾ Ayer, W. A.; Saeedi-Ghomi, M. H.; van Eggen, D.; Tagle, B.; Clardy, J. *Tetrahedron* **1981**, *37*, 379.

⁽¹¹⁾ Schmidt, J. A. R.; Lobkovsky, E. B.; Coates, G. W. J. Am. Chem. Soc. 2005, 127, 11426.

⁽¹²⁾ Cortez, G. S.; Oh, S. H.; Romo, D. Synthesis 2001, 11, 1731.

⁽¹³⁾ Annis, G. D.; Ley, S. V.; Self, C. R.; Sivaramakrishnan, R. J. Chem.

Soc., Perkin Trans. I 1. (14) MM2 calculations yield for the parent cis and trans systems steric energies of 34.1 and 68.7 kcal/mol, respectively. The calculations were carried out with CS Chem3D Pro.

in 1997,¹⁵ using the Rosenfeld method developed in the CADPAC program by Amos.¹⁶ Then, scientists reported various improved methods of optical rotation calculations.¹⁷ Now, it is popular to use B3LYP/aug-cc-pVDZ//B3LYP/6-31G(d) for prediction of the optical rotation in chiral rigid compounds. The computed optical rotation values for each enantiomer are summarized in Tables 2 and 3 of the Supporting Information. The sum of optical rotations for vibralactone (1) is -127.4° and for its enantiomer is $+127.4^{\circ}$. The first value is very close to the experimental value of -135.1° , which strongly suggests the configuration of vibralactone given in structure 1.

Surprisingly, initial attempts to proof the cis junction between the lactone and the cyclopentene ring in 1 by ROESY experiments showed no correlations between the protons at C5 and C8. From the calculations given in the Supporting Information, the distances between 5-H and the methylene protons 8a-H and 8b-H in the most stable cisconformer 1e are 3.1 and 3.9 Å, respectively. These distances decrease to 2.6 and 2.8 Å in the third most stable conformation 1b (relative energy of 0.322 kcal/mol) and the second most stable conformation 1c (0.241 kcal/mol), respectively.

Interestingly, a structurally closely related metabolite, percyquinnin (2), of undetermined relative and absolute configuration has recently been isolated from cultures of *Stereum complicatum*.¹⁸ 2 displays significant lipase-inhibiting properties. After completion of our work, Dr. M. Brönstrup, Aventis-Sanofi, kindly informed us that plane structure 2 for percyquinnin is wrong and has to be changed into that of vibralactone 1. The chiroptical properties of percyquinnin are still unknown.

Other bioactive β -lactones with a fused 4/5 ring system are salinosporamide A (**3**) and omuralide (**4**) (Figure 2).^{19–22} Salinosporamide A (**3**) and omuralide (**4**) are potent naturally derived substances which inhibit proteasome function with very high selectivity. Proteasome inhibition offers considerable promise in the therapy of a number of types of cancer and is already used for multiple myeloma.²³ A test on the potential proteasome activity of vibralactone (**1**) against chymotrypsin was planned.

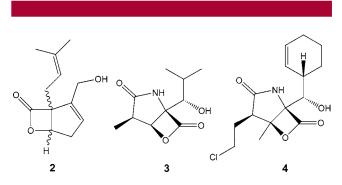


Figure 2. Structure of percyquinnin (2), salinosporamide A (3), and omuralide (4).

The structure of vibralactone (1) is interesting in connection with tetrahydrolipstatin (orlistat), a β -lactone-type natural lipase inhibitor. The pancreatic lipase activity of vibralactone (1) was measured using 4-methylmubelliferyl oleate (4-MU oleate) as a substrate. It showed pancreatic lipase inhibitory activity with IC₅₀ of 0.4 μ g/mL. The details are given in the Supporting Information. Tetrahydrolipstatin (orlistat), a reduced form of the natural product lipstatin, is an antiobesity agent marketed under the tradename Xenical that was recently approved by the FDA as the first over-the-counter weight-loss medication.²⁴ The β -lactone-containing natural products inhibit gastric and pancreatic lipase by blocking hydrolysis of triglycerides and thus the uptake of fatty acids from diet.²⁵ The mechanism of inhibition involves covalent but reversible modification of the active site serine via acylation by the β -lactone. The pancreatic lipase inhibitory activity of vibralactone (1) is probably due to the same mechanism.

Vibralactone (1): The culture broth was filtered to remove the mycelium. The filtrate (12 L) was then successively extracted twice with ethyl acetate, and the crude extract (1.75 g) was chromatographed on a silica gel column using a CHCl₃/MeOH gradient. Several fractions of increasing polarity were collected. Fraction II (120 mg) eluted with CHCl₃/MeOH (98:2, v/v) was further subjected to column chromatography over silica gel and Sephadex LH-20, using a petroleum ether/ethyl acetate (8:1, v/v) and CHCl₃/MeOH (1:1, v/v), respectively, to yield 65 mg of **1** as a colorless oil: $[\alpha]_D^{26} = -135.1$ (c = 0.52 in CHCl₃); IR (neat) $\nu =$ 3423 (OH), 2971, 2915, 2860, 1816 (β-lactone), 1112, 1000, 836 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, reference, $\delta = 7.26$ ppm) $\delta = 1.62$ (s, 3 H, 11-H), 1.71 (s, 3 H, 12-H), 2.41 (dd, J = 15.1, 7.3 Hz, 1 H, 8b-H), 2.60 (dd, J = 15.1, 7.3 Hz, 1 H, 8a-H), 2.71 (d, 1 H, J = 18.8 Hz, 4 α -H), 2.77 (dd, J =18.8, 4.3 Hz, 4 β -H), 4.22 (s, 2 H, 13-H), 4.79 (d, J = 4.3Hz, 1 H, 5-H), 5.11 (t, *J* = 7.3 Hz, 1 H, 9-H), 5.60 ppm (s, 1 H, 2-H); ¹³C NMR (125 MHz, CDCl₃, reference, $\delta = 77.0$ ppm) $\delta = 18.0 (C11), 25.8 (C12), 27.6 (C8), 37.3 (C4), 61.3$ (C13), 75.1 (C1), 78.5 (C5), 117.2 (C9), 122.4 (C2), 136.0

⁽¹⁵⁾ Polavarapu, P. L. Mol. Phys. 1997, 91, 551.

⁽¹⁶⁾ Amos, R. D. Chem. Phys. Lett. 1982, 87, 23

^{(17) (}a) Cheeseman, J. R.; Frisch, M. J.; Delvin, G. J.; Stephens, P. J. J. Phys. Chem. A 2000, 104, 1039. (b) Urbanova, M.; Setnicka, V.; Devlin, F. J.; Stephens, P. J. J. Am. Chem. Soc. 2005, 127, 6700. (c) Ruud, K.; Helgaker, T. Chem. Phys. Lett. 2002, 352, 533. (d) Ruud, K.; Zanasi, R. Angew. Chem., Int. Ed. 2005, 44, 3594. (e) Tam, M. C.; Russ, N. J.; Crawford, T. D. J. Chem. Phys. 2004, 121, 3550. (f) Crawford, T. D.; Owens, L. S.; Tam, M. C.; Schreiner, P. R.; Koch, H. J. Am. Chem. Soc. 2005, 127, 1368.

⁽¹⁸⁾ Hopmann, C.; Kurz, M.; Mueller, G.; Toti, L. (Aventis Pharma G.m.b.H), EP 1142886, 2001; *Chem. Abstr.* **2001**, 287595.

⁽¹⁹⁾ Feling, R. H.; Buchanan, G. O.; Mincer, T. J.; Kauffman, C. A.; Jensen, P. R.; Fenical, W. Angew. Chem., Int. Ed. 2003, 42, 355.

⁽²⁰⁾ Omura, S.; Fujimoto, T.; Otoguro, K.; Matsuzaki, K.; Moriguchi, R.; Tanaka, H.; Sasaki, Y. J. Antibiot. **1991**, 44, 113.

⁽²¹⁾ Omura, S.; Fujimoto, T.; Otoguro, K.; Matsuzaki, K.; Moriguchi, R.; Tanaka, H.; Sasaki, Y. J. Antibiot. **1991**, 44, 117.

⁽²²⁾ Corey, E. J.; Reichard, G. A.; Kania, R. Tetrahedron Lett. 1993, 34, 6977.

^{(23) (}a) Richardson, P. G.; Hideshima, T.; Anderson, K. C. *Cancer Control* **2003**, *10*, 361. (b) Steinberg, D. *Scientist* **2003**, *17* (S2), S18. (c) Adams, J. *Proteasome Inhibitors in cancer Therapy*; Human Press: New York, 2004.

⁽²⁴⁾ Ma, G.; Zancanella, M.; Oyola, Y.; Richardson, R. D.; Smith, J. W.; Romo, D. Org. Lett. **2006**, 8, 4497.

^{(25) (}a) Hadvery, P.; Lengsfeld, H.; Wolfer, H. Biochem. J. **1988**, 256, 357. (b) Borgstrom, B. Biochem. Biophys. Acta **1988**, 962, 308.

(C10), 146.6 (C3), 173.0 ppm (C7); MS (ESI⁺) m/z (%) 231 (100) [M + Na]⁺, 439 (85) [2M + Na]⁺; HR-ESI-MS m/z calcd for C₁₂H₁₆O₃Na [M + Na]⁺ 231.1004, found 231.0997.

In total, 24 conformations of **1** were obtained in the gas phase at the B3LYP/6-31G(d) level of theory. The obtained magnetic shielding magnitudes of ¹³C NMR were recorded at the B3LYP/6-311+G(2d,p) level and were corrected using the slope and intercept of the least-squares correlation line. All of the shieldings were finally converted into chemical shifts. The optical rotation values were calculated using B3LYP/aug-cc-pVDZ theory. The Boltzmann formula was used to produce the sum of six different conformational optical rotations. The details are given in the Supporting Information.

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Supporting Information Available: NMR data of **1**, computations, and bioassay experiment (PDF). This material is available free of charge via the Internet at http://pubs.acs.org.

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